

Prenylated isoflavonoids from *Millettia pervilleana*[☆]Giovanna Palazzino^{a,*}, Philippe Rasoanaivo^b, Elena Federici^a, Marcello Nicoletti^c,
Corrado Galeffi^a^aLaboratorio di Chimica del Farmaco, Istituto Superiore di Sanità, V. le Regina Elena 299, I-00161 Rome, Italy^bInstitut Malgache de Recherches Appliquées, Laboratoire de Phytochimie-Pharmacologie, B.P.3833, 101 Antananarivo, Madagascar^cDipartimento di Farmacologia delle Sostanze Naturali e Fisiologia Generale, Università "La Sapienza", P. le A. Moro 5, I-00185 Rome, Italy

Received 5 April 2002; received in revised form 25 September 2002

Abstract

From the root bark of *Millettia pervilleana*, which had shown significant cytotoxic activity, a 3-phenylcoumarin, named pervilleanine, two new pterocarpanes, pervilline and pervillinine, and one known, emoroidocarpan, were isolated in addition to rotenone and 3 α -hydroxyrotenone. The anticancer activity of two previously isolated isoflavanones, pervilleanone and 3'-*O*-demethylpervilleanone is reported.

© 2003 Elsevier Science Ltd. All rights reserved.

Keywords: *Millettia pervilleana*; Leguminosae; Coumarins; Pervilleanine; Pterocarpanes; Pervilline; Pervillinine; Cytotoxic and anti-cancer activity

1. Introduction

The genus *Millettia* (Leguminosae) includes about 150 subtropical species. Phytochemical research has revealed isoflavonoids as the main constituents of the genus.

Our previous study on the chloroform extract of root bark of *Millettia pervilleana* Viguier of Madagascar (IC₅₀ 0.12 $\mu\text{g ml}^{-1}$ on KB cells, Galeffi et al., 1997) resulted in the isolation of two prenylated isoflavanones, pervilleanone (**1**) and 3'-*O*-demethylpervilleanone (**2**) from fraction D endowed with mild cytotoxicity (IC₅₀ 0.57 $\mu\text{g ml}^{-1}$). The anticancer activity of **1** and **2**, performed by NCI on human tumor cell lines of lung, breast and CNS, is now reported (Table 1).

From two more active fractions of the same extract, further isoflavonoids, namely a 3-phenylcoumarin, pervilleanine (**3**), two new pterocarpanes, pervilline (**4**) and pervillinine (**5**), and one known pterocarpan, emor-

oidocarpan (**6**) (Machocho et al., 1995), besides rotenone and 3 α -hydroxyrotenone, were isolated.

2. Results and discussion

2.1. Characterization of the constituents

Fraction B (IC₅₀ 0.047 $\mu\text{g ml}^{-1}$ on KB cells) obtained from the chloroform extract of *M. pervilleana* (Galeffi et al., 1997) was submitted to counter-current distribution (CCD) and two main substances, **3** and **6**, were thus isolated. The less polar one, pervilleanine (**3**), C₂₂H₁₈O₆ (HR-EI-MS, *m/z* 378.1124 [M]⁺, calc. 378.1103), had an UV spectrum (see Section 3.2.1) showing extended conjugation. Its ¹H and ¹³C NMR spectra obtained by HETCOR and DEPT experiments showed the presence of a methylenedioxy group (δ_{H} 5.97, *s*) and an *ortho,ortho* disubstituted methoxy group (δ_{H} 3.54, δ_{C} 60.9). The fourth oxygen atom of the molecule belonged to a α,α -dimethylpyrane ring (δ_{H} 1.43, *s*, 2 Me; 5.69 and 6.89, both *d*, *J* = 10.2 Hz) originated by cyclisation of a prenyl group, whereas the last two oxygen atoms were those of a coumarin unit (δ_{CO} 163.6). The presence among the ¹³C NMR signals of **3** of an *ortho,ortho*-dioxxygen substituted oxygen-bearing carbon, δ_{C} 132.5, and of two *ortho* hydrogens, δ_{H} 7.55 and 6.71, *d*, *J* = 8.2 Hz (the latter shif-

[☆] Presented at the 10th Congresso Nazionale della Società Italiana di Fitochimica, Florence, Italy, 7–10 May 2000.

* Corresponding author. Tel.: +39-06-4990-2550; fax: +39-06-4938-7100.

E-mail address: palazzin@iss.it (G. Palazzino).

ted upfield as well the corresponding carbon, δ_{C} 113.1) accounted for a 2-methoxy-3,4-methylenedioxyphenyl ring linked in position 3 to the coumarin system.

Two of the three remaining aromatic hydrogens, δ_{H} 6.92, *d*, and 6.86, *d*, in the *peri* position, at C-4 and C-5, were mutually coupled ($J=1.7$ Hz) whereas the last hydrogen, δ_{H} 6.84, *s*, was attached to C-8, δ_{C} 108.2, which is *ortho,ortho*-dioxxygen substituted. The structure of pervilleanine, **3**, was thus fully established.

Substance **6**, $\text{C}_{21}\text{H}_{18}\text{O}_5$ (HR–EI–MS, m/z 350.1093 $[\text{M}]^+$, calc. 350.1154), was emoroidocarp, a pterocarp isolated from *Tephrosia emroides* A. Rich. (Machoch et al., 1995). The complete assignment of its ^{13}C NMR frequencies is reported in Table 2. The

absolute *R* configuration of both chiral centres 6a and 11a (which was earlier not reported, as well the specific optical rotation) can be assigned on the basis of *cis*-diaxial relationship of their hydrogens ($J=6.8$ Hz) and the negative Cotton effect of the CD curve (Baruah et al., 1984).

Fraction C (IC_{50} 0.08 $\mu\text{g ml}^{-1}$ on KB cells) was likewise submitted to CCD and column chromatography and four substances were isolated in order of increasing polarity, two new pterocarps, pervilline (**4**) and pervillinine (**5**), rotenone and 3 α -hydroxyrotenone.

Pervilline (**4**), $\text{C}_{21}\text{H}_{20}\text{O}_5$ (HR–EI–MS, m/z 352.1263 $[\text{M}]^+$, calc. 352.1311), was a pterocarp as demonstrated by the sequence $\text{O}-\text{CH}_2-\text{CH}(\text{Ar})-\text{CH}-\text{O}$ (CH_2 ,

Table 1
3-Cell line assay for the primary anticancer screening of **1** and **2** at conc. 10^{-4} M

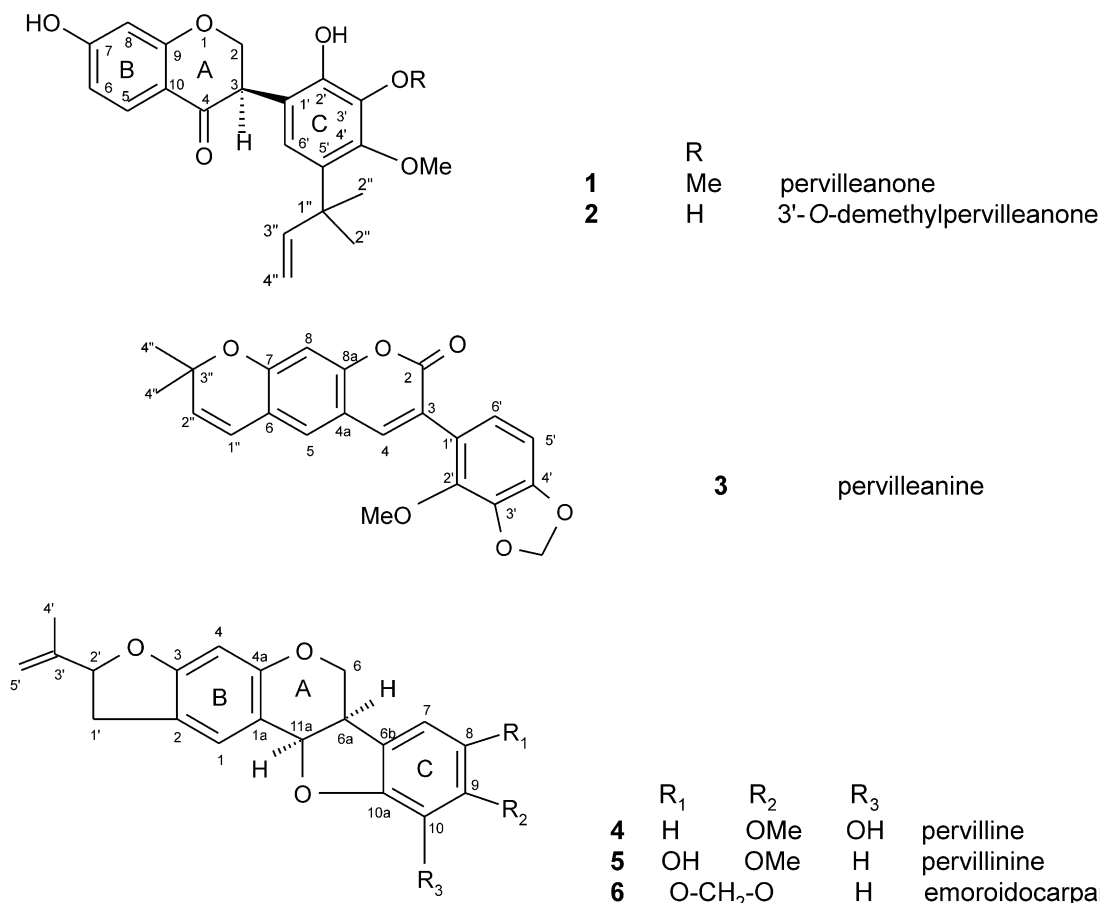
Compound	Growth percentages of the treated cell lines		
	(Lung) NCI-H460	(Breast) MCF7	(CNS) SF-268
Pervilleanone, 1	–36	–38	–27
3'-O-Demethylpervilleanone, 2	–2	–56	–42

Negative values of the growth percentage correspond to $T_i < T_z$, where T_z is the cell population at time zero and T_i is the cell population in the presence of the drug after 48 h.

Table 2
 ^{13}C and ^1H NMR spectral data of pterocarps **4**, **5** and **6**

Position 4 (CDCl_3)		5 (CDCl_3)		6 (CDCl_3)	
	δ_{C} δ_{H} (<i>J</i> in Hz)		δ_{C} δ_{H} (<i>J</i> in Hz)		δ_{C} δ_{H} (<i>J</i> in Hz)
1	126.8 7.33, <i>s</i>	126.3 7.33, <i>s</i>		126.2 7.20, <i>s</i>	
1a	111.7	n.o.		111.8	
2	120.7	120.8		120.8	
3	161.2	161.2		161.4	
4	98.1 6.35, <i>s</i>	98.2 6.35, <i>s</i>		98.2 6.38, <i>s</i>	
4a	156.0	156.1		156.3	
6	66.5 3.60, <i>dd</i> (10.8; 10.6); 4.19, <i>dd</i> (10.8; 4.9)	66.6 3.60, <i>t</i> (11.0); 4.19, <i>dd</i> (11.0; 5.0)		66.5 3.55, <i>t</i> (11.4); 4.16, <i>dd</i> (11.4; 5.1)	
6a	40.1 3.50, <i>m</i> (10.6; 6.6; 4.9)	40.3 3.50, <i>m</i> (11.0; 6.7; 5.0)		40.1 3.40, <i>m</i> (11.4; 6.8; 5.1)	
6b	121.7	118.1		118.1*	
7	114.7 6.72, <i>d</i> (8.0)	110.4 6.79, <i>s</i>		104.7 6.67, <i>s</i>	
8	103.6 6.42, <i>d</i> (8.0)	139.9		141.8	
9	148.0	146.9		147.5	
10	130.6	94.7 6.41, <i>s</i>		93.7* 6.33, <i>s</i>	
10a	146.1	152.8		154.1*	
11a	79.8 5.52, <i>d</i> (6.6)	78.7 5.52, <i>d</i> (6.7)		79.0 5.40, <i>d</i> (6.8)	
1'	33.8 2.97, <i>dd</i> (15.8; 7.6); 3.26, <i>dd</i> (15.8; 9.8)	33.9 2.97, <i>dd</i> (15.7; 7.7); 3.26, <i>dd</i> (15.7; 9.8)		33.8 2.94, <i>dd</i> (15.6; 8.0); 3.23, <i>dd</i> (15.6; 9.8)	
2'	86.6 5.16, <i>dd</i> (9.8; 7.6)	86.7 5.16, <i>dd</i> (9.8; 7.7)		86.6 5.12, <i>dd</i> (9.8; 8.0)	
3'	143.8	143.8		143.7	
4'	17.1 1.76, <i>s</i>	17.2 1.76, <i>s</i>		17.1 1.70, <i>s</i>	
5'	112.1 5.05; 4.88, <i>br s</i>	112.1 5.05; 4.88, <i>br s</i>		112.1 5.01; 4.85, <i>d</i> (1.1)	
OMe	56.4 3.84, <i>s</i>	56.2 3.84, <i>s</i>			
OCH ₂ O				101.2 5.83; 5.89, <i>d</i> (2.5)	

* The assignments of Machoch et al. (1995) for these carbons are not correct.



δ_{H} 3.60, *dd*, $J=10.8$; 10.6 Hz, and 4.19, *dd*, $J=10.8$; 4.9 Hz; CH-Ar, 3.50, *m*, $J=10.6$; 6.6; 4.9 Hz; CH-O 5.52, *d*, $J=6.6$ Hz). The substitution of the benzofuran ring C was shown by the COLOC connectivities of C-10a (δ_{C} 146.1) and C-9 (δ_{C} 148.0) with the hydrogen at δ_{H} 6.72 (*d*, $J=8.0$ Hz) and by the connectivity of C-6b (δ_{C} 121.7) with the hydrogen at δ_{H} 6.42 (*d*, $J=8.0$ Hz). These two *ortho* hydrogens are in position 7 and 8, respectively. The connectivity of C-9 with the methoxy group at δ_{H} 3.84 and the ^{13}C resonance of the latter (δ_{C} 56.4) accounted for its mono-*ortho* substituted position 9. The remaining hydroxy group was thus located in position 10 in agreement with the upfield resonance of C-10 (δ_{C} 130.6) due to the *ortho,ortho*-dioxxygen substitution.

Moreover in **4** a prenyl group was linked to ring B to give a methylethenyldihydrofuran-fused ring (H₂-1', δ_{H} 2.97, *dd*, and 3.26, *dd*; H-2', 5.16, *dd*; H-4', 1.76, *s*; H₂-5', 4.88 and 5.05, *br s*). The latter is located as in emoroidocarpan in agreement with the presence of an *ortho,ortho*-dioxxygen substituted methine (δ_{H} 6.35, *s*, δ_{C} 98.1) in position 4.

The absolute configuration of **4** is identical to that of emoroidocarpan on account of the coupling constant $J_{6a/11a}$ (6.6 Hz) and the negative Cotton effect.

Pervillinine (**5**), C₂₁H₂₀O₅ (HR-EI-MS, m/z 352.1251 [M]⁺, calc. 352.1311), differed from pervilline on the

substitution in ring C (see Table 2). In the former, the two hydrogens at δ_{H} 6.79, *s* and δ_{H} 6.41, *s* were located in the *para* positions 7 and 10, respectively, and the methoxy group (δ_{H} 3.84) in position 9 inasmuch its irradiation by NOE experiments enhanced the signal of the more shielded H-10 (δ_{H} 6.41). The hydroxy group was thus located in position 8.

2.2. In vitro anticancer activity

Isoflavonoids have shown activity in the search for antiviral and antineoplastic or cancer chemopreventive agents of plant origin. Thus, some isoflavonoids from genus *Millettia* showed inhibitory effects against Epstein–Barr virus (Ito et al., 2000), whereas those of *M. pachycarpa* inhibited the activities of murine retroviral reverse transcriptase and human DNA-polymerases (Ono et al., 1989).

Rotenone and 3 α -hydroxyrotenone, which were found in the cytotoxic fraction C of *M. pervilleana*, showed inhibition of TPA-induced ornithine decarboxylase at the level of its mRNA expression and were therefore regarded as promising cancer chemopreventive agents (Gerhauser et al., 1995).

Pervilleanone (**1**) and 3'-O-demethylpervilleanone (**2**), isolated from fraction D of *M. pervilleana* endowed with mild cytotoxicity, showed growth inhibition of the

human cancer cell lines of lung, breast and CNS (Table 1) used in the NCI antitumor prescreen.

3. Experimental

3.1. General

A Craig-Post apparatus (200 stages, 10:10 ml, upper and lower phase) was used for the separation by CCD. TLC: silica gel F₂₅₄, cyclohexane–EtOAc (1:1). CD: Jasco 710. ¹H and ¹³C NMR: 500.13 and 125.77 MHz, respectively, Bruker AM 500. Chemical shifts are given in δ (ppm) from internal TMS. EI–MS: 70 eV, HP 5989A. HR–EI–MS: VG 7070 EQ–HF.

The bioassay tests on lung, breast and CNS cells were performed by the NCI, Bethesda (USA).

3.2. Separation

Fraction B (1.4 g) was submitted to CCD with solvent system H₂O–Me₂CO–EtOH–cyclohexane (2:6:2:8). Two chromatographically pure fractions, pervilleanine (3, 39 mg) and emoroidocarpan (6, 68 mg) were thus obtained.

Fraction C (1.4 g) was submitted to CCD with solvent system H₂O–Me₂CO–EtOH–cyclohexane (5:4:4:10) and four fractions were obtained. Each fraction was subsequently purified by column chromatography on silica gel 60 (70–230 mesh), eluent cyclohexane–EtOAc (8:2) and thus pervilline (4, 26 mg), pervillinine (5, 31 mg), rotenone (89 mg) and 3 α -hydroxyrotenone (56 mg) were obtained in order of increasing polarity. The last two substances were identified by comparison of their physico-chemical data.

3.2.1. Pervilleanine (3)

Pale crystals from *n*-hexane, mp 177–179 °C. UV (MeOH): λ_{\max} nm (log ϵ): 324 (4.09), 239 (4.02). ¹H NMR (CDCl₃): δ 1.43 (6H, *s*, 2 Me-4''); 3.54 (3H, *s*, OMe); 5.69 (1H, *d*, *J* = 10.2 Hz, H-2''); 5.97 (2H, *s*, O–CH₂–O); 6.71 (1H, *d*, *J* = 8.2 Hz, H-5'); 6.84 (1H, *s*, H-8); 6.86 (1H, *d*, *J* = 1.7 Hz, H-5); 6.89 (1H, *d*, *J* = 10.2 Hz, H-1''); 6.92 (1H, *d*, *J* = 1.7 Hz, H-4); 7.55 (1H, *d*, *J* = 8.2 Hz, H-6'). ¹³C NMR (CDCl₃): δ 28.0 (2 Me, C-4''); 60.9 (OMe); 77.4 (C-3''); 101.2 (O–CH₂–O); 108.1 (C-4a); 108.2 (C-8); 108.8 (C-6); 110.9 (C-1'); 111.4 (C-4); 113.1 (C-5'); 115.4 (C-1''); 123.8 (C-6'); 124.7 (C-5); 125.9 (C-3); 130.3 (C-2''); 132.5 (C-3'); 147.5 (C-7); 147.6 (C-4'); 148.4 (C-2'); 156.2 (C-8a); 163.6 (C-2). EI–MS, *m/z* (rel. int.): 378 ([M]⁺, 44), 363 (100), 320 (17), 187 (13), 159 (15), 145 (10). Molecular formula C₂₂H₁₈O₆, HR–EI–MS, *m/z*: 378.1124 [M]⁺, calc. 378.1103.

3.2.2. Pervilline (4)

White powder from *n*-hexane, mp 112–114 °C. [α]_D²⁰ –192° (CHCl₃; *c* 0.12). CD: [θ]₂₄₀ –44 600 (MeOH; *c* 0.013). UV (MeOH): λ_{\max} nm (log ϵ): 298 (3.9), 220

(4.1). ¹H and ¹³C NMR data in Table 2. EI–MS, *m/z* (rel. int.): 352 ([M]⁺, 22), 350 (24), 337 (22), 162 (37), 149 (100). Molecular formula C₂₁H₂₀O₅, HR–EI–MS, *m/z*: 352.1263 [M]⁺, calc. 352.1311.

3.2.3. Pervillinine (5)

Yellow powder from *n*-hexane, mp 152–154 °C. [α]_D²⁰ –189° (CHCl₃; *c* 0.07). CD: [θ]₂₄₀ –46 800 (MeOH; *c* 0.011). UV (MeOH): λ_{\max} nm (log ϵ): 298 (3.9), 220 (4.1). ¹H and ¹³C NMR data in Table 2. EI–MS, *m/z* (rel. int.): 352 ([M]⁺, 100), 337 (58), 322 (18). Molecular formula C₂₁H₂₀O₅, HR–EI–MS, *m/z*: 352.1251 [M]⁺, calc. 352.1311.

3.2.4. Emoroidocarpan (6)

White crystals from *n*-hexane, mp 189–190 °C. [α]_D²⁰ –268° (CHCl₃; *c* 0.11). CD: [θ]₂₄₀ –45 200 (MeOH; *c* 0.012). UV (MeOH): λ_{\max} nm (log ϵ): 300 (2.9), 220 (3.9). ¹H and ¹³C NMR data in Table 2. EI–MS, *m/z* (rel. int.): 350 ([M]⁺, 100), 335 (87). Molecular formula C₂₁H₁₈O₅, HR–EI–MS, *m/z*: 350.1093 [M]⁺, calc. 350.1154.

Acknowledgements

The authors are greatly indebted to the National Cancer Institute, Bethesda (USA), for the anticancer assays.

References

- Baruah, P., Barua, N.C., Sharma, R.P., Baruah, J.N., Kulanthaivel, P., Herz, W., 1984. Flavonoids from *Millettia pulchra*. *Phytochemistry* 23, 443–447.
- Galeffi, C., Rasoanaivo, P., Federici, E., Palazzino, G., Nicoletti, M., Rasolondratovo, B., 1997. Two prenylated isoflavanones from *Millettia pervilleana*. *Phytochemistry* 45, 189–192.
- Gerhauser, C., Mar, W., Lee, S.K., Suh, N., Luo, Y., Kosmeder, J., Luyengi, L., Fong, H.H.S., King-Horn, A.D., Moriarty, R.M., Mehta, R.G., Constantinou, A., Moon, R.C., Pezzuto, J.M., 1995. Rotenoids mediate potent cancer chemopreventive activity through transcriptional regulation of ornithine decarboxylase. *Nature Medicine* 1, 260–266.
- Ito, C., Itoigawa, M., Tan, H.T.W., Tokuda, H., Mou, X.Y., Mukainaka, T., Ishikawa, T., Nishino, H., Furukawa, H., 2000. Antitumor-promoting effects of isoflavanoids on Epstein-Barr virus activation and two-stage mouse skin carcinogenesis. *Cancer Letters* 152, 187–192.
- Machocho, A.K., Lwande, W., Jondiko, J.I., Moreka, L.V.C., Hassanal, A., 1995. Three new flavonoids from the root of *Tephrosia emoroides* and their antifedant activity against the larvae of the spotted stalk borer *Chilo partellus* Swinhoe. *International Journal of Pharmacognosy* 33, 222–227.
- Ono, K., Nakane, H., Meng, Z.M., Ose, Y., Sakai, Y., Mizuno, M., 1989. Differential inhibitory effects of various herb extracts on the activities of reverse transcriptase and various deoxyribonucleic acid (DNA) polymerase. *Chemical and Pharmaceutical Bulletin* 37, 1810–1812.